

1.161. This allowed assignment of the signals at δ 1.161 and 2.422 to C21-H and C20-H, respectively. This means that the remaining highly deshielded dimethyl singlet (δ 1.100) must be caused by C28-H and C29-H and the dimethyl doublet (δ 0.933) by C26-H and C27-H.

24,24-Dimethyl-5 α -cholesta-7,25-dien-22-yn-3 β -ol (6) acetate: mp 138–140 °C; RRT(GC) 1.50, RRT(HPLC) 0.37; MS, m/z (assignment, relative intensity) 450.3461 (C₃₁H₄₆O₂, M⁺, 34, calcd 450.3494), 435.3247 (C₃₀H₄₃O₂, 12), 407.2922 (C₂₈H₃₉O₂, 4), 475.3064 (C₂₈H₃₉, 4), 367.2634 (C₂₅H₃₅O₂, 4), 315.2286 (C₂₁H₃₁O₂, 18), 313.2114 (C₂₁H₂₉O₂, 10), 299.2046 (C₂₀H₂₇O₂, 6), 267.2071 (C₂₀H₂₇, 2), 255.2102 (C₁₉H₂₇, 38), 253.1945 (C₁₉H₂₅, 6), 239.1843 (C₁₈H₂₃, 6), 229.0000 (C₁₇H₂₅, 11), 213.1657 (C₁₆H₂₁, 9), 43.0193 (C₂H₃O₁, 100). Hydrogenation of 6-acetate (4 h) afforded a mixture of the acetates of 7 (70%) and 10 (30%), which was separated by HPLC. The MS and ¹H NMR data of these acetates were indistinguishable from those of the hydrogenation products of 5-acetate.

Decoupling experiments: Irradiation of the methyl doublet at δ 1.170 collapsed the signal at δ 2.468 (1 H, qd) into a doublet (J = 6.8) and irradiation at δ 2.468 collapsed the methyl doublet at δ 1.170. Thus we found C20-H and C22-H.

(22Z)-24,24-Dimethyl-5 α -cholesta-7,22-dien-3 β -ol (7) acetate: mp 195–197 °C; RRT(GC) 1.88, RRT(HPLC) 0.90; MS, m/z (assignment, relative intensity) 454.3804 (C₃₁H₅₀O₂, M⁺, 54), 439.3545 (C₃₀H₄₇O₂, 18), 411.3237 (C₂₈H₄₃O₂, 18), 394.3556 (C₂₉H₄₆, 8), 351.3080 (C₂₆H₃₉, 9), 342.2587 (C₂₃H₃₄O₂, 14), 315.2308 (C₂₁H₃₁O₂, 30), 313.2131 (C₂₁H₂₉O₂, 75), 299.2018 (C₂₀H₂₇O₂, 7), 288.2065 (C₁₉H₂₈O₂, 20), 273.1856 (C₁₈H₂₅O₂, 8), 255.2081 (C₁₉H₂₇, 67), 253.1962 (C₁₉H₂₅, 7), 241.1967 (C₁₈H₂₅, 9), 229.1937 (C₁₇H₂₅, 37), 213.1684 (C₁₆H₂₁, 20), 81.0717 (C₆H₉, 100).

Decoupling experiments: Irradiation at δ 0.983 collapsed the methine signal at δ 2.653 (qdd) into a double doublet (J = 9.6,

7.4), whereas irradiation at δ 2.653 collapsed the methyl doublet at δ 0.983 into a singlet and the methine signal at δ 4.987 (dd) into a doublet (J = 12.5). Further irradiation at δ 4.987 (1 H, dd) collapsed the methine signal at δ 2.653 (qdd) into a quadruple doublet (J = 6.6, 9.4) and the methine doublet at δ 5.053 into a singlet. On the basis of these results the signals at δ 0.983, 2.653, 4.987, and 5.053 were assigned to C21-H, C20-H, C22-H, and C23-H, respectively.

24,24-Dimethyl-5 α -cholest-7-en-3 β -ol (10) acetate: mp 180–183 °C; RRT(GC) 2.02, RRT(HPLC) 1.22; MS, m/z (relative intensity) 456 (M⁺, 100), 441 (18), 413 (3), 396 (59), 381 (12), 315 (11), 288 (8), 273 (9), 255 (67), 229 (20), 213 (29). ¹H NMR data were reported earlier.^{22a} The previously unreported ¹³C NMR data are included in Table I. Assignment of the side-chain ¹³C signals was made with the aid of ¹³C NMR data for the relevant model paraffins.³⁹

Acknowledgment. We thank Drs. Y. Fujimoto (The Institute of Physical and Chemical Research, Saitama, Japan), K. Furuhashi (Institute of Applied Microbiology, The University of Tokyo), N. Shimizu (Hitachi Chemical Co., Ibaraki, Japan), and T. Takido (Nihon University) for NMR spectra and Dr. M. Aimi (Nihon University) for the mass spectra.

Registry No. 1, 118112-67-7; 1 acetate, 118112-68-8; 2, 118203-79-5; 4, 82467-98-9; 5, 118112-69-9; 5 acetate, 118112-73-5; 6, 118142-17-9; 6 acetate, 118112-71-3; 7, 118112-70-2; 7 acetate, 118112-72-4; 10, 105097-81-2; 10 acetate, 105097-84-5.

(39) Lindeman, L. P.; Adams, J. Q. *Anal. Chem.* 1971, 43, 1245.

C-Glycosides. 7.[†] Stereospecific C-Glycosylation of Aromatic and Heterocyclic Rings

S. Czernecki* and G. Ville[†]

Laboratoire de Chimie des Glucides, Université P. et M. Curie, Tour 54-55, E1, 4 Pl. Jussieu, 75005 Paris, France

Received July 25, 1988

Stereospecific C-glycosylation of aromatic and heterocyclic rings can be realized by reacting the corresponding organolithium derivatives with benzylated lactones and reducing the so-obtained lactols with triethylsilane in the presence of boron trifluoride etherate at low temperature. Debenzylation proceeds without opening of the ring in pyrano series, but with opening in furano series.

Because of their biological importance, considerable effort has been devoted toward the synthesis of C-glycosides during recent years.¹

With the goal of devising new and efficient methodologies for the stereocontrolled synthesis of this class of compounds, we are studying the reactivity of several sugar derivatives, diversely activated at the anomeric carbon atom, with a variety of organometallic reagents. Several combinations have already been studied and the results published: glycals with arylpalladium species,² peracylated enones³ and protected 1,2-anhydro sugars^{4a} with organocuprates.

We report herein our results concerning the reaction of organolithium derivatives with protected lactones and the reduction of the products so obtained into C-glycosides (Scheme I).

Sugar lactones have previously been employed for that purpose by Kishi et al.⁵ to prepare allyl-C-glycosides. In their approach to C-nucleosides, Asbun and Binkley⁶ and Ogura et al.⁷ effected C–C bond formation in this fashion but were not successful in reduction of the lactol product.

(1) For a recent review, see the special issue of *Carbohydr. Res.* on C-glycoside synthesis: *Carbohydr. Res.* 1987, 171.

(2) Czernecki, S.; Dechavanne, V. *Can. J. Chem.* 1983, 61(3), 533.

(3) Bellosta, V.; Czernecki, S. *Carbohydr. Res.* 1987, 171, 279.

(4) (a) Bellosta, V.; Czernecki, S. *J. Chem. Soc., Chem. Commun.*, in press. (b) Bellosta, V. Thèse, Université P. et M. Curie, Paris, 1987.

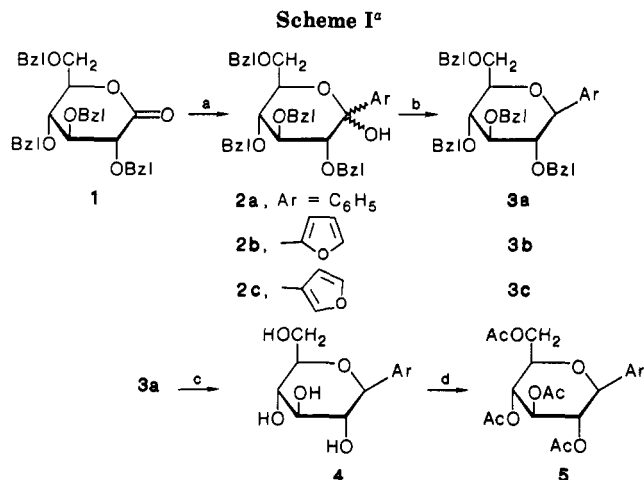
(5) Lewis, M. D.; Cha, J. K.; Kishi, Y. *J. Am. Chem. Soc.* 1982, 104, 4976.

(6) Asbun, W.; Binkley, S. B. *J. Org. Chem.* 1968, 33, 140.

(7) Ogura, H.; Takahashi, H. *J. Org. Chem.* 1974, 39, 1374.

[†]Communicated in part at the 3rd European Symposium on Carbohydrates, Grenoble, France, 1985. For preceding paper, see ref 3.

[†]Current address: Laboratoire de Chimie Organique et Cinétique, Université de Picardie, 33 rue Saint Leu, 80000 Amiens, France.



^aKey: (a) ArLi (1.05 equiv), THF, -78 °C; (b) Et₃SiH/BF₃, Et₂O/MeCN/-40 °C→room temperature; (c) 10% Pd on C/MeOH/H₂; (d) Ac₂O/pyridine.

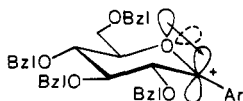
Since very reactive organometallic reagents were employed, stable protective groups (benzyl) were necessary.

2,3,4,6-Tetra-*O*-benzyl-D-glucopyranolactone⁸ (1) was used as a model compound for examining the stereochemical outcome of the reduction step and verifying that the deprotection by hydrogenolysis would not cleave the O₅-C₁ bond in the resulting C-glycoside 3.⁹

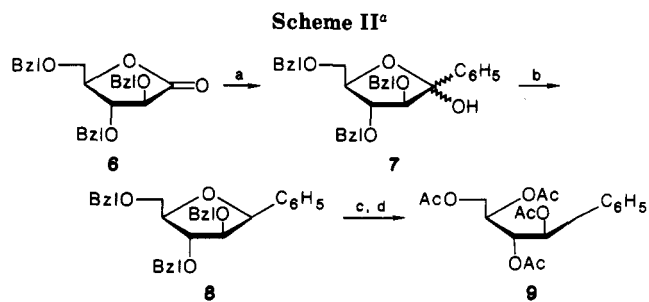
Reaction of 1 with 1 equiv of phenyllithium in THF (-78 → 20 °C) afforded an α/β mixture of (2,3,4,6-tetra-*O*-benzyl-1-*C*-phenyl)-D-glucopyranose (2a) in 85% yield. The reduction of the hemiacetal function of 2a with triethylsilane in the presence of boron trifluoride etherate¹⁰ (acetonitrile at -40 °C for 1 h) was stereospecific, and the C-glycoside 3a was isolated in 80% yield. Assignment of stereochemistry was made by examining of the physical constants and spin-spin coupling constants for the ring protons in the 250 MHz ¹H NMR spectrum of the tetraacetate 5, obtained by debenzoylation (H₂/Pd-C/MeOH/room temperature) followed by acetylation (Ac₂O/pyridine). The physical constants of 5 were in agreement with the literature data^{4b} for the β isomer including the value of 9.7 Hz for $J_{1,2}$.

In order to verify the stereospecificity of the reduction step and the absence of the α isomer, hydrogenolysis and acetylation were carried out, without any purification, on crude 3a. VPC analysis of crude 5 showed only one peak, the retention time of which was identical with that of an authentic sample available in our laboratory.^{4b} A mixed injection with the α anomer^{4b} gave two well-resolved peaks (see Experimental Section).

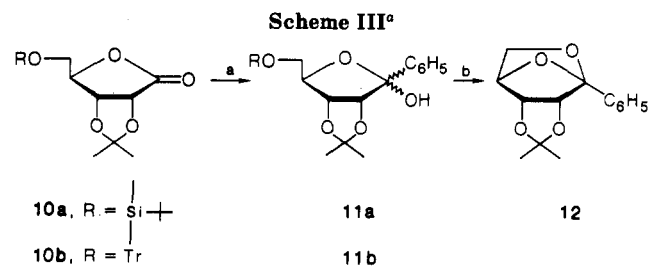
The stereospecificity of the reduction may be due to stabilization of the carbenium intermediate by the anomeric effect, favoring hydride attack at the α face.¹¹



An advantage of our procedure is the fact that the organolithium reagent can be prepared by exchange from the



^aSee Scheme I.



^aSee Scheme I.

aromatic ring or the corresponding halide allowing a regioselective C-glycosylation. This was demonstrated by C-glycosylation of furan at the 2- and 3-positions by starting respectively from furan and 3-bromofuran. 3-Furyl-C-glycoside subunits are often present in natural compounds, and some of them exhibit interesting biological activities.¹² 2-Furyl-C-glycosides could be useful intermediates in the synthesis of base-modified nucleosides, because the furan ring can be converted into a large number of other heterocyclic systems.¹³

Reduction of the furan adducts proceeded smoothly at low temperature, affording only one isomer 3b or 3c, whose anomeric configurations were also found to be β , since $J_{1,2}$ was 9.7 Hz for 3b and 10.0 Hz for 3c. An anomeric mixture of furyl-C-glycosides was obtained by Maeba et al.¹⁴ by reaction of 2,3,5-tri-*O*-benzyl-D-ribofuranosyl bromide with 2-(chloromercurio)furan.

The extension of this methodology for glycosylation of aromatic rings with pentofuranosyl moieties was also studied since it would provide a new route to C-nucleosides when applied to heterocycles.

The same sequence of reactions was performed on *O*-benzyl-protected arabinolactone 6 (Scheme II).

When submitted to catalytic hydrogenation and acetylation, the intermediate 8¹⁵ was transformed into 1,2,3,4-tetraacetoxy-5-phenyl-(2*R*,3*S*,4*R*)-pentane (9). The presence of four singlets ($\delta \approx 2$ ppm, 12 H) and of an ABX multiplet for two benzylic hydrogens in the ¹H NMR spectrum of 9 clearly indicated that the O₄-C₁ bond was cleaved during deprotection. This reduction cleavage of the carbohydrate ring could not be avoided by using other solvents (AcOEt or THF).¹⁶

In order to circumvent the problem of ring opening in the furano series, other blocking groups, stable to organolithium reagents, were evaluated with the commercially

(12) (a) Kawakami, Y.; Nagai, Y.; Nezu, Y.; Sato, T.; Kumi, T.; Kagei, K. *Chem. Pharm. Bull.* 1987, 35, 4839. (b) Harde, C.; Bohlman, F. *Tetrahedron* 1988, 44(1), 81.

(13) Van der Plas, H. C. *Ring Transformations of Heterocycles*; Academic Press: New York, 1973; Vol. 1, p 151.

(14) Maeba, I.; Iwata, K.; Usami, F.; Furukawa, H. *J. Org. Chem.* 1983, 48, 2998.

(15) One isomer was isolated in 85% yield, but the anomeric configuration was not firmly established; it is written as β for convenience of reading.

(16) Hacksell, U.; Daves, G. D., Jr. *J. Org. Chem.* 1983, 48, 4144.

(8) Kuzuhara, H.; Fletcher, H. G., Jr. *J. Org. Chem.* 1967, 32, 2531.

(9) While this paper was in preparation, a similar approach appeared: Kraus, G. A.; Molina, M. T. *J. Org. Chem.* 1988, 53, 752.

(10) Fraïnnet, E.; Esclamadon, C. C. R. *Hebd. Seances Acad. Sci.* 1962, 254, 1814.

(11) *Anomeric Effect: Origin and Consequences*; Szarek, W. A., Horton, D., Eds.; American Chemical Society: Washington, DC, 1979; ACS Symposium Series 87.

available ribonolactone (Scheme III).

Reaction of **10a** and **10b** with phenyllithium afforded the hemiacetals **11a** and **11b** (90% and 81% yield, respectively). Reduction of these compounds as above led to the same product **12**, the structure of which was determined by spectroscopic methods. In this case, with acid-labile protective groups at C-5, a cyclization is possible, and it took place faster than the reduction. Such a propensity to form 1,5-anhydro derivatives from ribofuranosyl compounds has been already observed.¹⁷

The procedure described herein constitutes a very convenient method for the C-glycosylation of aromatic and heterocyclic rings with pyranosyl moieties. Further studies with other protective groups are currently in progress in our laboratory to extend it to the furano series.

Experimental Section

General Methods. Optical rotations were measured on a Perkin-Elmer 141 polarimeter. IR spectra (film, KBr disk) were recorded on a Unicam SP3-300 spectrophotometer. ¹H NMR spectra were recorded in CDCl₃ or C₆D₆, using Me₄Si as internal standard, at 90 MHz with a Varian EM-390 instrument or at 250 MHz with a Bruker apparatus operating in the FT mode. Gas chromatographic analysis was carried out on a Girdel 75 FD 2 instrument equipped with flame-ionization detectors and fitted with a 1-m 3% w/w phenyldiethanolamine succinate (PDEAS) on Chromosorb W AW DMCS column at 155 °C. Analytical TLC was performed on precoated alumina plates (E. Merck silica gel 60F₂₅₄). For flash chromatography, E. Merck silica gel 60 (230–400 mesh) and anhydrous solvents were used. Mass spectra (MS) were obtained on a Nermag R10-10 spectrometer using chemical ionization (NH₃).

General Procedure. (2,3,4,6-Tetra-*O*-acetyl-β-D-glucopyranosyl)benzene (**5**). The following preparation illustrates the typical procedure.

Arylation. Lactone **1**⁸ (1.08 g, 2 mmol) was dissolved in 5 mL of THF, and the mixture was cooled to -78 °C under argon. Phenyllithium (1.25 mL of 1.7 M, 2.1 mmol) was then added, and the mixture was stirred for 2 h at -78 °C. The temperature was then allowed to rise to room temperature while the mixture was further stirred for 2 h. After hydrolysis, extraction with ether, and drying over anhydrous MgSO₄, the solvent was evaporated under reduced pressure, yielding 1.056 g (85.7% yield) of **2a** as a colorless oil, pure by TLC (*R*_f 0.45 with ether-pentane, 2:1): IR 3400 (br), 3090, 3060, 3030, 2970, 2920, 2860, 1595, 1450, 1360, 1210, 1090, 1025, 910, 735, and 695 cm⁻¹.

Reduction. To a solution of 616 mg (1 mmol) of **2a** in 10 mL of acetonitrile, cooled to -40 °C, was added 320 μL (2 molar equiv) of triethylsilane and 140 μL (1 molar equiv) of BF₃·Et₂O. After the solution had been stirred at -40 °C for 1 h, 1 mL of a saturated aqueous solution of K₂CO₃ was added, and the mixture was stirred for 1 h at room temperature. The aqueous layer was extracted with ether, and the organic phase was dried over anhydrous Mg₂SO₄. Evaporation of the solvent afforded 540 mg of a yellowish oil, which was purified by column chromatography on silica (pentane-ether), yielding 480 mg (0.8 mmol; 80% yield) of pure **3a**: [α]_D²⁰ 29.4° (c 1.5, CHCl₃); IR 3090, 3060, 3030, 2900, 2860, 1600, 1580, 1490, 1450, 1360, 1210, 1090, 1065, 1030, 1000, 910, 735, and 695 cm⁻¹. Anal. Calcd for C₄₀H₄₀O₅: C, 79.97; H, 6.71. Found: C, 79.59; H, 6.82.

Debzylation. Compound **3a** (120 mg, 0.2 mmol) was dissolved in 5 mL of methanol, and 20 mg of 10% palladium on charcoal was added. The mixture was vigorously stirred in a hydrogenation apparatus. After completion of the reaction (6 h), the catalyst was filtered off and the solvent was evaporated, yielding 44 mg (0.18 mmol; 90% yield) of chromatographically homogeneous product **4a** (*R*_f 0.22 with AcOEt): mp 49–50 °C; [α]_D²⁰ 30.3° (c 1.9, CH₃OH); ¹H NMR [(CD₃)₂SO] δ 3.17 (dd, 1 H, H-2, *J*_{1,2} = 9.3 Hz, *J*_{2,3} = 10.1 Hz), 3.20–3.29 (m, 4 H, H-3, H-4,

H-5, OH), 3.46 (dd, 1 H, H-6, *J*_{5,6} = 5.4 Hz, *J*_{6,6'} = 11.6 Hz), 3.70 (dd + br s, 2 H, H-6', OH, *J*_{5,6'} = 1.3 Hz), 4.01 (d, 1 H, H-1), 4.83 (br s, 3 H, OH), 7.06–7.30 (m, 5 H, aromatic H).

Acetylation. The hydroxyl groups were acetylated by using a classical procedure (Ac₂O-pyridine), giving 62 mg (0.15 mmol; 84% yield) of pure **5**. The crude product was analyzed by VPC on a 5% PDEAS on 45–50 mesh Chromosorb W NAW column (1 m × 0.375 in.) at 155 °C. The retention time of the observed peak was 23.8 min, identical with that of an authentic sample available in our laboratory.^{4b} A mixed injection with the α isomer gave two well-resolved peaks (retention time of α: 19.8 min): [α]_D²⁰ -20° (c 1.1, CHCl₃); mp 156–157 °C. Anal. Calcd for C₂₀H₂₄O₉: C, 58.82; H, 5.88. Found: C, 58.88; H, 5.84.

2'-(2,3,4,6-Tetra-*O*-benzyl-β-D-glucopyranosyl)furan (3b**)** was obtained in 77% yield (454 mg) by the same sequence, by condensing of 2-furyllithium¹⁸ with **1** (540 mg; 1 mmol) and was characterized as follows: mp 111 °C; [α]_D²⁰ 10.42°¹⁹ (c 1.2, CH₂Cl₂); MS, *m/z* (relative abundance) 591 (M + 1, 45), 561 (M - HCO, 100); ¹H NMR (C₆D₆) δ 3.47 (ddd, 1 H, H-5, *J*_{4,5} = 9.6 Hz, *J*_{5,6} = 3.8 Hz, *J*_{5,6'} = 5.6 Hz), 3.56 (m, 2 H, H-6, H-6'), 3.62–3.95 (m, 2 H, H-3, H-4), 3.97 (t, 1 H, H-2, *J*_{1,2} = 9.7 Hz, *J*_{2,3} = 9.2 Hz), 4.10–5.00 (8 d, 8 H, benzylic CH₂), 4.30 (d, 1 H, H-1), 6.05 (dd, 1 H, H-4', *J*_{3,4'} = 3.2 Hz, *J*_{4,5'} = 1.8 Hz), 6.25 (dd, 1 H, H-3'), *J*_{3,5'} = 0.7 Hz), 7.05–7.37 (m, 21 H, aromatic H and H-5'). Anal. Calcd for C₃₈H₃₈O₆: C, 77.29; H, 6.44. Found: C, 77.32; H, 6.54.

3'-(2,3,4,6-Tetra-*O*-benzyl-β-D-glucopyranosyl)furan (3c**)** was similarly obtained in 30% yield (177 mg) by condensation of 3-furyllithium²⁰ with **1** (540 mg; 1 mmol) and characterized as follows: mp 140–141 °C; [α]_D²⁰ -9.4° (c 0.1, CHCl₃); MS, *m/z* (relative abundance) 591 (M + 1, 4), 561 (M - HCO, 100); ¹H NMR (C₆D₆) δ 3.21 (dd, 1 H, H-2, *J*_{1,2} = 10 Hz, *J*_{2,3} = 9.3 Hz), 3.30–3.42 (m, 2 H, H-6, H-6', *J*_{6,6'} = 11.6 Hz), 3.43 (dd, 1 H, H-4, *J*_{3,4} = 9.3 Hz, *J*_{4,5} = 10 Hz), 4.02 (t, 1 H, H-3), 4.07 (m, 1 H, H-5), 4.50–5.02 (8 d, 8 H, benzylic H), 4.96 (d, 1 H, H-1), 5.26 (d, 1 H, H-4', *J*_{4,5'} = 3.4 Hz), 7.16–7.20 (m, 2 H, H-2', H-5'), 7.28–7.40 (m, 20 H, aromatic H). Anal. Calcd for C₃₈H₃₈O₆: C, 77.29; H, 6.44. Found: C, 77.18; H, 6.31.

1,2,3,4-Tetraacetoxy-5-phenyl-(2*R*,3*S*,4*R*)-pentane (9**)** was obtained from 2,3,5-tri-*O*-benzyl-D-arabinofuranolactone (**6**)²¹ (836 mg; 2 mmol) in 58% overall yield (306 mg) by using the phenylation/reduction/hydrogenolysis/acetylation sequence already described for the preparation of **5** and was characterized as follows: mp 106 °C; [α]_D²⁰ 43.7° (c 1.2, CHCl₃); ¹H NMR (CCl₄) δ 1.95, 2.02, 2.04, 2.18 (4 s, 12 H, 4 CH₃CO), 2.76 (dd, 1 H, H-5, *J*_{5,5'} = 13.5 Hz, *J*_{4,5} = 8.1 Hz), 2.83 (dd, 1 H, H-5', *J*_{4,5'} = 5.3 Hz), 4.12 (dd, 1 H, H-1, *J*_{1,2} = 5.1 Hz, *J*_{1,1'} = 12.5 Hz), 4.23 (dd, 1 H, H-1', *J*_{1,2} = 2.8 Hz), 5.14 (ddd, 1 H, H-2, *J*_{2,3} = 8 Hz), 5.32 (dd, 1 H, H-3, *J*_{3,4} = 2.6 Hz), 5.39 (dd, 1 H, H-4), 7.08–7.27 (m, 5 H, aromatic H). Anal. Calcd for C₁₉H₂₄O₈: C, 59.99; H, 6.36. Found: C, 60.00; H, 6.32.

1,5-Anhydro-2,3-*O*-isopropylidene-1-*C*-phenyl-β-D-ribofuranose (12**)** was obtained from 5-*O*-(*tert*-butyldimethylsilyl)-2,3-*O*-isopropylidene-D-ribofuranolactone²² (**10a**) (604 mg; 2 mmol) by using the same phenylation/reduction sequence in 51% overall yield (253 mg) and characterized as follows: mp 118 °C; [α]_D²⁰ -87.9° (c 1.4, CHCl₃); MS, *m/z* (relative abundance) 249 (M + 1, 100), 105 (M - PhCO, 70); ¹H NMR (CCl₄) δ 1.22, 1.42 (2 s, 6 H, 2 CH₃), 3.46 (d, 1 H, H-2, *J*_{2,3} = 7.5 Hz), 3.60 (dd, 1 H, H-3, *J*_{3,4} = 3.7 Hz), 4.29, 4.44 (2 d, 2 H, H-5, H-5', *J*_{5,5'} = 5.5 Hz, *J*_{4,5} = *J*_{4,5'} = 0 Hz), 4.75 (d, 1 H, H-4), 7.40–7.55 (m, 5 H, aromatic H). Anal. Calcd for C₁₄H₁₆O₄: C, 67.73; H, 6.50. Found: C, 67.68; H, 6.54.

Preparation of **12 from 5-*O*-trityl-2,3-*O*-isopropylidene-D-ribofuranolactone (**10b**)²³** (860 mg; 2 mmol) was carried out by using the same technique, in 36% overall yield (178 mg).

(18) Ramanathan, V.; Levine, R. *J. Org. Chem.* **1962**, *27*, 1216.

(19) Grynkiewicz, G.; BeMiller, J. N. *Carbohydr. Res.* **1984**, *131*, 273. The differences in physical constants of **3b** between our results and those reported in this paper, mp 94 °C and [α]_D²⁰ 71° (c 1, CH₂Cl₂), are likely due to an incorrect assignment of anomeric configuration by the authors.

(20) Gilman, H.; Melstrom, D. S. *J. Am. Chem. Soc.* **1948**, *70*, 1655.

(21) Rabinsohn, Y.; Fletcher, H. G., Jr. *J. Org. Chem.* **1967**, *32*, 3452.

(22) Cheng, J. C.; Hacksell, U.; Daves, G. D., Jr. *J. Org. Chem.* **1985**, *50*, 2778.

(23) Camps, P.; Cardellach, J.; Font, J.; Ortuño, R. M.; Ponsati, O. *Tetrahedron* **1982**, *38*(15), 2395.

(17) (a) Barker, G. R.; Lock, M. V. *J. Chem. Soc.* **1950**, 23. (b) Vis, E.; Fletcher, H. G., Jr. *J. Am. Chem. Soc.* **1957**, *79*, 1182. (c) Kraska, B.; Lichtentaler, F. W. *Chem. Ber.* **1981**, *114*, 1636. (d) Izquierdo Cubero, I.; Garcia Poza, D. *Carbohydr. Res.* **1985**, *138*, 139.